Day 4 Worksheet – Read mapping and visualization

Author: Qing Yang, 2021
Edited: Lynn Sanford, 2022

1. Make sure you have the following files in your …**/day4/trimmomatic/** directory:

```
26M Jul 13 14:31 chr21Eric_repA.RNA.end1.trimmed.fastq
  0 Jul 13 14:31 chr21Eric_repA.RNA.end1.unpaired.fastq
26M Jul 13 14:31 chr21Eric_repA.RNA.end2.trimmed.fastq
  0 Jul 13 14:31 chr21Eric_repA.RNA.end2.unpaired.fastq
```

2. Create new directory (***mkdir***) named **hisat2**, under …**/day4/**, for the output directory for mapped reads.

```
-bash-4.2$ mkdir hisat2
mkdir: created directory ‘hisat2’
-bash-4.2$ ls -lsh
total 1.0K
  0 drwxrwsr-x+ 2 qiya9811 dowelldegrp 0 Jul 13 14:24 fastq/
  0 drwxrwsr-x+ 2 qiya9811 dowelldegrp 0 Jul 13 14:33 fastqc/
  0 drwxrwsr-x+ 2 qiya9811 dowelldegrp 0 Jul 13 14:33 hisat2/
512 drwxrwsr-x+ 2 qiya9811 dowelldegrp 1 Jul 13 14:27 scripts/
512 drwxrwsr-x+ 2 qiya9811 dowelldegrp 4 Jul 13 14:31 trimmomatic/
```

3. Transfer the d4_mapping script from the sread2022 scripts directory to your scripts directory.

```
[lynn-sanford@ip-172-31-18-92 day4]$ rsync /scratch/Shares/public/sread2022/scripts/
day4/d4_mapping.sbatch /scratch/Users/lynn-sanford/day4/
[lynn-sanford@ip-172-31-18-92 day4]$ ls scripts/
d4_mapping.sbatch  d4_trim_qc.sbatch
```

4. Edit the new “d4_mapping.sbatch” using the text editor **vim**. First, edit the SBATCH configuration to meet the needs of read mapping:

a. Change the name of the job to something more useful, such as “hisat2_mapping”.
b. Replace <EMAIL> with your own email address to which you want to receive any notifications.
c. Replace <USERNAME> with your own username to complete the path directory to where to store the error and output files.
d. Complete the following fields: nnodes, ntasks, mem and time. Hisat2 can use multiple processors per input file. So, 1 node, 8 tasks/processors/CPUs, 2 Gb for memory and 90 minutes for wall-time should be enough.

```
#!/bin/bash
#SBATCH --job-name=d4_mapping                          # Job name
#SBATCH --mail-type=ALL                          # Mail events (NONE, BEG
#SBATCH --mail-user=<YOUR_EMAIL_HERE>            # Where to send mail
#SBATCH --nodes=1                                # Number of nodes reques
#SBATCH --ntasks=8                               # Number of CPUs (proces
#SBATCH --mem=2gb                                # Memory limit
#SBATCH --time=01:30:00                          # Time limit hrs:min:sec
#SBATCH --partition=short                        # Partition/queue reques
#SBATCH --output=/scratch/Users/<USERNAME>/day4/eofiles/%x.%j.out
laced with job_name and the %j by the job id
#SBATCH --error=/scratch/Users/<USERNAME>/day4/eofiles/%x.%j.err
```

5. Next, assign path variables. In this case, we will specify two directories, both under **DATADIR**. **TRIM** stores the directory path to trimmed reads. **HISAT2** stores the directory path to output mapped reads.

```
### Assigns path variables

DATADIR=/scratch/Users/<USERNAME>/day4/
HISAT2=${DATADIR}/hisat2
TRIM=${DATADIR}/trimmomatic
```

6. Next, load the modules/software needed for mapping reads and file conversion:

```
### Loads modules
module load hisat2/2.1.0
module load samtools/1.8
```

7. And finally, specify the read mapping and file conversion commands. Note that you could instead break up the command onto many lines using the character "\" at the end of every line. These \ characters are ignored by the computer, but will help you identify each part of the command more easily:

NOTE: The genome index is located at **/scratch/Shares/public/genomes/hisatfiles/hg38/HISAT2/genome**

```
### <SOFTWARE SPECIFICS>

## Map trimmed reads to reference genome

hisat2 --very-fast -x /scratch/Shares/public/genomes/hisatfiles/hg38/HISAT2/genome \
-1 ${TRIM}/chr21Eric_repA.RNA.end1.trimmed.fastq \
-2 ${TRIM}/chr21Eric_repA.RNA.end2.trimmed.fastq \
> ${HISAT2}/chr21Eric_repA.RNA.sam \
2> ${HISAT2}/chr21Eric_repA.hisat2_maptstats.txt

## Convert mapped reads to sorted bam file
### convert SAM to BAM
samtools view -@ 8 -bS -o ${HISAT2}/chr21Eric_repA.RNA.bam \
${HISAT2}/chr21Eric_repA.RNA.sam

### sort bam file
samtools sort -@ 8 ${HISAT2}/chr21Eric_repA.RNA.bam \
> ${HISAT2}/chr21Eric_repA.RNA.sorted.bam

### index sorted bam file
samtools index ${HISAT2}/chr21Eric_repA.RNA.sorted.bam \
${HISAT2}/chr21Eric_repA.RNA.sorted.bam.bai
```

8. Before you close vim, make sure to save your edits by press Esc button to exit insertion mode, then type in ***:wq*** to save and quit vim.

9. Now that the job script is complete, submit the job by type in ***sbatch*** command. While waiting for the job to execute, you can check the job status using the command ***squeue -u <USERNAME>***:

```
-bash-4.2$ sbatch mapping.sbatch
Submitted batch job 7730124
-bash-4.2$ squeue -u qiya9811
             JOBID PARTITION     NAME     USER ST       TIME  NODES NODELIST(REASON)
           7730124     short hisat2_m qiya9811  R       0:07      1 fijinode-12
```

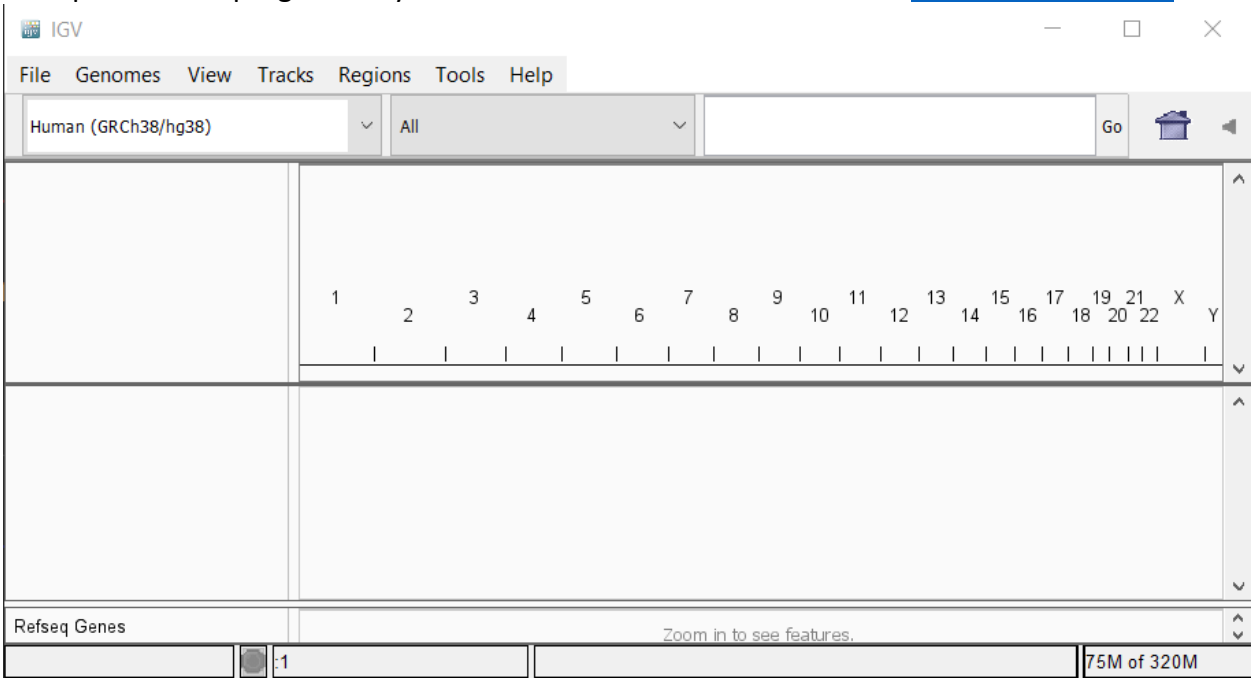
10. Finally, check the output directory …/day4/hisat2/ - there should be 5 different files:

```
-bash-4.2$ ls -lsh
total 172M
1.0K -rw-rw-r--+ 1 qiya9811 dowelldegrp  613 Jul 13 16:33 chr21Eric_repA.hisat2_maptstats.txt
 24M -rw-rw-r--+ 1 qiya9811 dowelldegrp  24M Jul 13 16:33 chr21Eric_repA.RNA.bam
127M -rw-rw-r--+ 1 qiya9811 dowelldegrp 127M Jul 13 16:33 chr21Eric_repA.RNA.sam
 19M -rw-rw-r--+ 1 qiya9811 dowelldegrp  19M Jul 13 16:33 chr21Eric_repA.RNA.sorted.bam
1.7M -rw-rw-r--+ 1 qiya9811 dowelldegrp 1.7M Jul 13 16:33 chr21Eric_repA.RNA.sorted.bam.bai
```

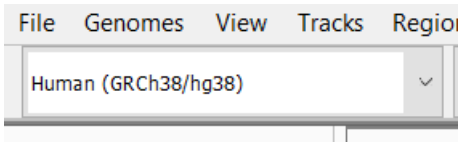
11. To visualize the mapped reads using IGV, you will need to transfer the sorted.bam and sorted.bam.bai files to your local machine. **rsync** the files from the AWS using a terminal on your local machine. Note that here, I've navigated to the directory for my desktop before rsyncing (Windows machine).

```
lsanford@DESKTOP-3GP5MRN:/mnt/c/Users/lsanford/Desktop$ rsync lynn-sanford@3.136.149.251:
/scratch/Users/lynn-sanford/day4/hisat2/chr21Eric_repA.RNA.sorted* ./
```

12. Open the IGV program on your local machine or in the browser at https://igv.org/app/



13. Before you load your bam files, make sure the genome is consistent with the reference genome that was used for read mapping. In this case, human hg38 genome is used. In a case where you need to load your custom genome, refer to the IGV manual page: https://software.broadinstitute.org/software/igv/LoadGenome



14. Finally, load the bam files we just created using HISAT2 and samtools by clicking on "**File**" > "**Load From File**", then choose the desired bam files in your HISAT2 output directory. To save time, we only mapped reads from chromosome 21 to the human genome, so to visualize the mapped reads, make sure to zoom into specific loci on chromosome 21. For example, the IFNAR1 gene is shown in the screenshot below. You can zoom in by typing in the gene name in the genome coordinate box, and by adjusting using the +/- button at the top right corner. Here, you should see the mapped reads summarized in both coverage and alignment formats: